

GnRH agonist 與 GnRH antagonist在卵巢顆粒細胞內的不同作用機轉

盧道權 醫師

在試管嬰兒的誘導排卵過程中，GnRH agonist與GnRH antagonist已經被廣泛使用來防止 premature LH surge。目前知道GnRH agonist是經由造成腦下垂體 desensitization 和 GnRH receptor down-regulation而達到抑制性腺刺激素 (gonadotropin)分泌的目的。GnRH antagonist則是和體內的 GnRH 直接競爭位在 gonadotropic cell membrane上的 receptor，一旦 antagonist 佔據了 receptor，血中性腺刺激素分泌濃度就明顯下降。近年來有愈來愈多的研究證明GnRH agonist與GnRH antagonist在不同的身體組織細胞也有不同的生理作用機轉。我們已經知道下視丘和腦下垂體是GnRH的主要來源及作用目標組織，然而許多研究指出下視丘外的GnRH及腦下垂體外的GnRH receptor，存在於生殖系統組織細胞，例如卵巢、子宮內膜組織、胎盤及子宮內膜癌細胞等。Ortmann and Diedrich 在1999年時發現很多腦下垂體外有GnRH-1 receptor的表現；也有人在人類卵巢(Kakar et al., 1992; Minaretzis et al., 1995; Peng et al., 1994)、黃體組織(Bramley et al., 1987)、顆粒黃體細胞(Brus et al., 1997)、排卵前的卵泡顆粒細胞層上(Choi et al., 2006)找到GnRH-1 receptor的mRNA。也有證據顯示GnRH在卵巢內會表現自我調控 (autocrine) 及周邊調控 (paracrine) 的行為去調控卵泡的成長(follicle development)和荷爾蒙的生成(steroidogenesis)(Andreu et al., 1998; Hsueh and Jones, 1981)。Lin et al. 在1999年已提到GnRH agonist在卵巢內的desensitization作用是不同於GnRH antagonist。很多研究也發現接受GnRH antagonist治療的女性，其打hCG當天的血清中 estradiol 濃度(Albano et al., 2000; Olivennes et al., 2000; Roulier et al., 2003)和卵泡液中 estradiol 濃度(Garcia-Velasco et al., 2001)遠比接受GnRH agonist治療者低許多(Figure 1)。所以這就說明了GnRH agonist與GnRH antagonist在卵巢內存在不同的作用機轉。這也是為什麼用GnRH antagonist比較可以預防卵巢過度反應症候群的原因(Hsieh et al., 2008; Lin et al., 2007)。

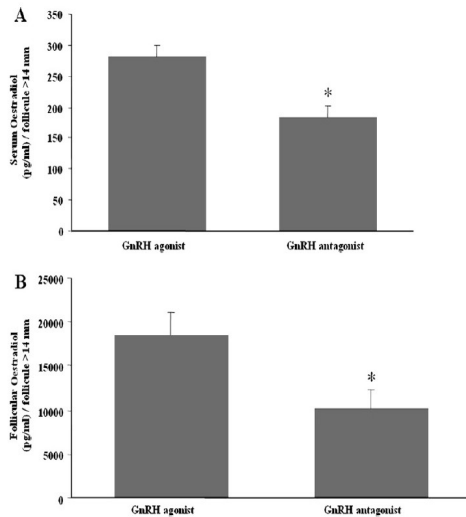


Figure 1 Concentrations of oestradiol in (A) serum and (B) follicular fluid (follicles >14 mm) in women undergoing gonadotrophin-releasing hormone (GnRH) agonist (n = 28) or antagonist (n = 22). Oestradiol was determined by radioimmunoassay. Values are mean \pm SD. *P < 0.05.

Annie 2010 年便探討了為何 GnRH agonist 與 GnRH antagonist protocol 的 estradiol 濃度有別的原因：首先卵巢顆粒細胞 (granulosa cell) 內的 estradiol 合成是來自於 aromatase enzyme expression 和 cAMP (30–50 adenosine monophosphate) pathway (Detail as figure below)。而使用 GnRH antagonist protocol 這一組病人的卵巢顆粒黃體細胞之 aromatase activity 和 aromatase (CYP19) gene expression 有顯著意義比較低 (Figure 2)。Garcia-Velasco 2001 年也提到 GnRH antagonist 組的卵泡液中之 estradiol : testosterone ratio 也是比較低。換言之使用 GnRH antagonist 這一組病人的 estradiol 濃度比較低是由於卵巢顆粒細胞內合成不足，不是鞘細胞 (theca cell) 提供的 androgenic precursors 不足。

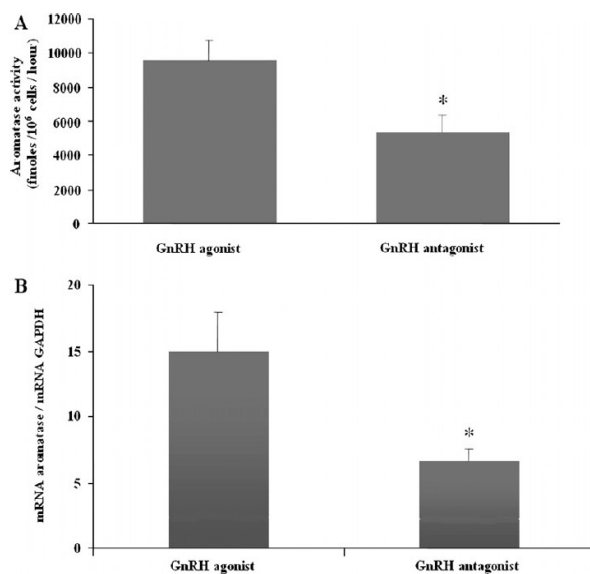
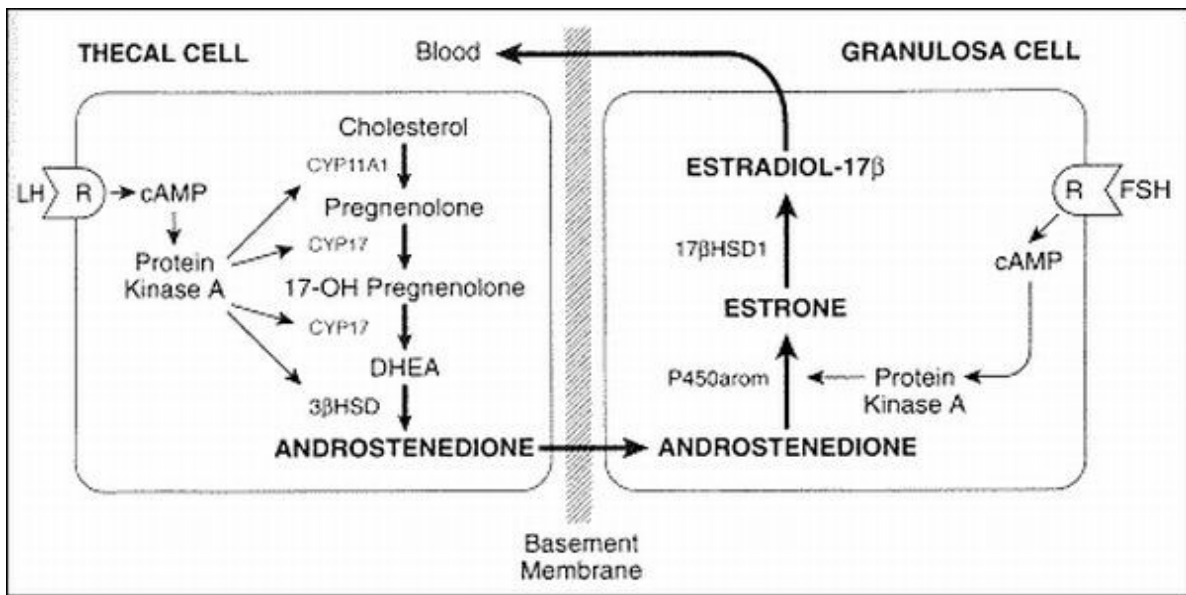
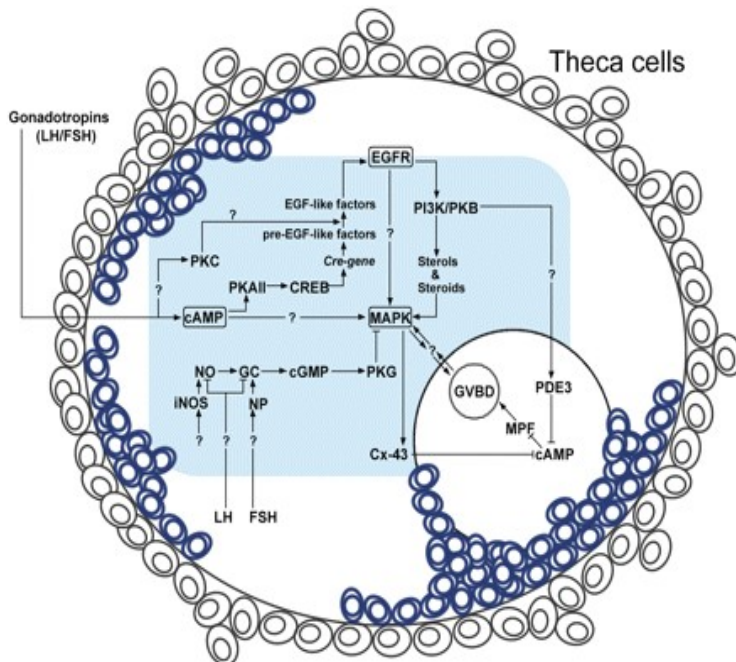
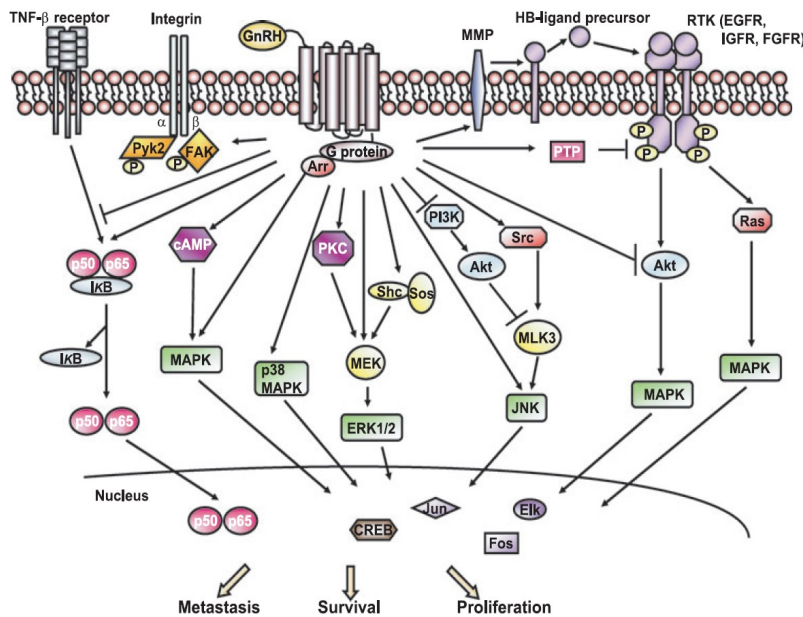


Figure 2 (A) Aromatase activity and (B) aromatase gene expression in human granulosa lutein cells treated with gonadotrophin-releasing hormone (GnRH) agonist or antagonist. Aromatase activity was quantified by $^3\text{H}_2\text{O}$ method. mRNA aromatase were quantified by real-time reverse-transcription PCR and values were normalized to mRNA GAPDH. Values are means \pm SD. *P < 0.05.



人類卵巢的GnRH-1 receptor比腦下垂體少 200 倍 (Minaretzis et al., 1995)。GnRH-1 receptor 經由活化 Gq/G11 heterotrimeric 蛋白而激活一系列的 mitogen-activated protein kinase 活動 (Figure 3)。





卵巢上的GnRH-1 receptor並沒有因為使用GnRH agonist或GnRH antagonist後減少。GnRH antagonist像在腦下垂體般透過直接競爭GnRH-1 receptor而達到減少 estradiol 合成；而GnRH agonist則是經由抑制PKC (protein kinase C) activity(Figure 4) 使得 estradiol 濃度比較高，因為活化PKC會抑制 gonadotrophin-induced steroidogenesis。

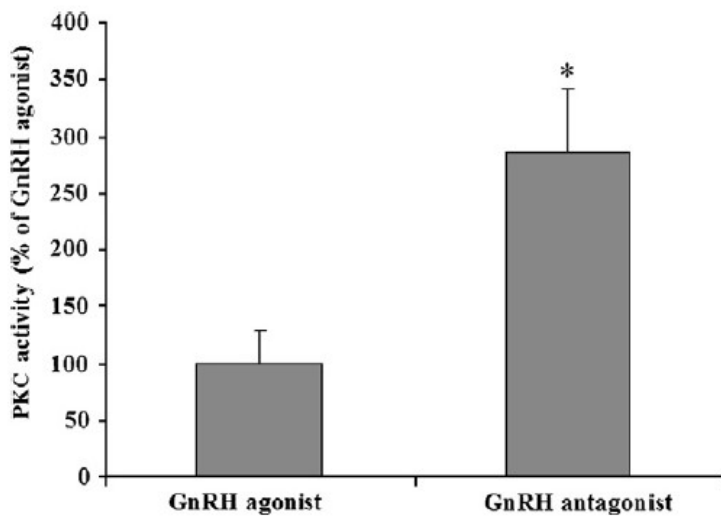


Figure 4 PKC activity in granulosa lutein cells treated with gonadotrophin-releasing hormone (GnRH) agonist or antagonist.

所以從目前有限的文獻資料中已經知道GnRH agonist與GnRH antagonist在卵巢顆粒細胞內有著不同的生理作用機轉，而這也還需要更多的研究去探討証實GnRH的生理、分子及細胞功能機轉。由於GnRH agonist與GnRH antagonist已廣泛的應用於臨床治療，如果我們能更清楚的了解GnRH的整個系統全貌，相信對於未來GnRH的臨床應用，會帶來更適合且更有效的治療。

參考文獻：

1. Annie B., Mohamad K., Herve' M., et al., 2010. GnRH agonist and GnRH antagonist protocols in ovarian stimulation: differential regulation pathway of aromatase expression in human granulosa cells. *Reprod. Biomed. Online* 21, 56–65.
2. Albano, C., Felberbaum, R.E., Smitz, J., et al., 2000. Ovarian stimulation with HMG: results of a prospective randomized phase III European study comparing the luteinizing hormone releasing hormone (LHRH)-antagonist cetrorelix and the LHRHagonist buserelein. European Cetrorelix Study Group. *Hum. Reprod.* 15, 526–531.
3. Andreu, C., Parborell, F., Vanzulli, S., et al., 1998. Regulation of follicular luteinization by a gonadotropin-releasing hormone agonist: relationship between steroidogenesis and apoptosis. *Mol. Reprod. Dev.* 51, 287–294
4. Bramley, T.A., Stirling, D., Swanston, I.A., et al., 1987. Specific binding sites for gonadotrophin-releasing hormone, LH/chorionic gonadotrophin, low-density lipoprotein, prolactin and FSH in homogenates of human corpus luteum. II: concentrations throughout the luteal phase of the menstrual cycle and early pregnancy. *J. Endocrinol.* 113, 317–327.
5. Brus, L., Lambalk, C.B., de Koning, J., et al., 1997. Specific gonadotrophin-releasing hormone analogue binding predominantly in human luteinized follicular aspirates and not in human pre-ovulatory follicles. *Hum. Reprod.* 12, 769–773.
6. Choi, J.H., Gilks, C.B., Auersperg, N., et al., 2006. Immunolocalization of gonadotropin-releasing hormone (GnRH)-I, GnRH-II, and type I GnRH receptor during follicular development in the human ovary. *J. Clin. Endocrinol. Metab.* 91, 4562–4570.
7. Garcia-Velasco, J.A., Isaza, V., Vidal, C., et al., 2001. Human ovarian steroid secretion in vivo: effects of GnRH agonist versus antagonist (cetrorelix). *Hum. Reprod.* 16, 2533–2539.
8. Hsieh, Y.Y., Chang, C.C., Tsai, H.D., 2008. Comparisons of different dosages of gonadotropin-releasing hormone (GnRH) antagonist, short-acting form and single, half-dose, long-acting form of GnRH agonist during controlled ovarian hyperstimulation and in vitro fertilization. *Taiwan. J. Obstet. Gynecol.* 47, 66–74.
9. Hsueh, A.J., Jones, P.B., 1981. Extrapituitary actions of gonadotropin-releasing hormone. *Endocr. Rev.* 2, 437–461.
10. Kakar, S.S., Musgrove, L.C., Devor, D.C., et al., 1992. Cloning, sequencing, and expression of human gonadotropin releasing hormone (GnRH) receptor. *Biochem. Biophys. Res. Commun.* 189, 289–295.
11. Lin, Y., Kahn, J.A., Hillensjo, T., 1999. Is there a difference in the function of granulosa-luteal cells in patients undergoing in-vitro fertilization either with gonadotrophin-releasing hormone agonist or gonadotrophin-releasing hormone antagonist? *Hum. Reprod.* 14, 885–888.
12. Lin, Y.H., Seow, K.M., Hsieh, B.C., et al., 2007. Application of GnRH antagonist in

combination with clomiphene citrate and hMG for patients with exaggerated ovarian response in previous IVF/ICSI cycles. *J. Assist. Reprod. Genet.* 24, 331–336.

13. Minaretzis, D., Jakubowski, M., Mortola, J.F., et al., 1995. Gonadotropin-releasing hormone receptor gene expression in human ovary and granulosa-lutein cells. *J. Clin. Endocrinol. Metab.* 80, 430–434.

14. Olivennes, F., Belaisch-Allart, J., Empeaire, J.C., et al., 2000. Prospective, randomized, controlled study of in vitro fertilization-embryo transfer with a single dose of a luteinizing hormone-releasing hormone (LH-RH) antagonist (cetorelix) or a depot formula of an LH-RH agonist (triptorelin). *Fertil. Steril.* 73, 314–320.

15. Ortman, O., Diedrich, K., Ortman and Diedrich 1999. Pituitary and extrapituitary actions of gonadotrophin-releasing hormone and its analogues. *Hum. Reprod.* 14 (Suppl. 1), 194–206.

16. Peng, C., Fan, N.C., Ligier, M., et al., 1994. Expression and regulation of gonadotropin-releasing hormone (GnRH) and GnRH receptor messenger ribonucleic acids in human granulosa-luteal cells. *Endocrinology* 135, 1740–1746.

17. Roulier, R., Chabert-Orsini, V., Sitri, M.C., et al., 2003. Depot GnRH agonist versus the single dose GnRH antagonist regimen (cetorelix, 3 mg) in patients undergoing assisted reproduction treatment. *Reprod. Biomed. Online* 7, 185–189.